

Quantification of the Polydisperse Heparin Analog Dociparstat by Signature Disaccharide Analysis in the Presence of Concomitant Enoxaparin Dosing in Support of a Phase 2/3 Study in Patients with Severe COVID-19



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OVERVIEW

Analysis of a polydisperse heparin analog therapeutic (CX-01, Dociparstat) was conducted in human plasma by signature disaccharide (NS6S) analysis, with NS6S being unique to CX-01 in native human plasma but common to the concomitantly dosed low molecular weight heparin (LMWH), Enoxaparin, in COVID-19 patients, demonstrating a novel quantification approach to concomitant related glycosaminoglycan (GAG) therapeutics. Signature disaccharide response correction was required to achieve accurate Dociparstat quantification in concomitantly dosed samples by in-run monitoring of an alternate signature disaccharide unique to the LMWH. The method was used to determine CX-01 concentrations in a Phase 2/3 study to evaluate the safety and efficacy of Dociparstat sodium for the treatment of severe COVID-19 in adults at high risk of respiratory failure.

INTRODUCTION

Bioanalytical methodologies were previously developed, extensively optimized, and validated at Q² Solutions for quantification of the novel CX-01 therapeutic GAG in rat, dog and human plasmas. The core methodology was based on concepts from the literature works of Linhardt¹, et al., focusing on the analysis of disaccharides derived from GAG macromolecule hydrolysis.

Key Optimization Aspects for Successful CX-01 Method Validations:

- GAG hydrolysis, signature disaccharide selectivity confirmation, disaccharide derivatization, HRMS detection with stable label ISTD disaccharide (NS6S-¹³C₆)

Key Development Aspects for CX-01 Analysis in the Presence of Enoxaparin:

- Response correction in the previously validated CX-01 method for "added" NS6S detected from patients receiving concomitant CX-01 and Enoxaparin treatments, where both treatment materials contain NS6S disaccharide subunits

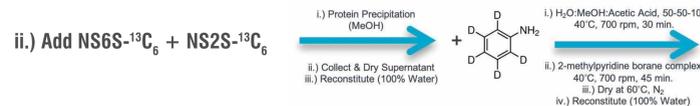
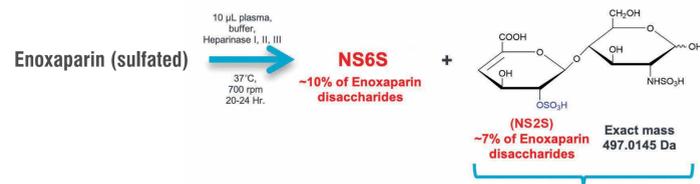
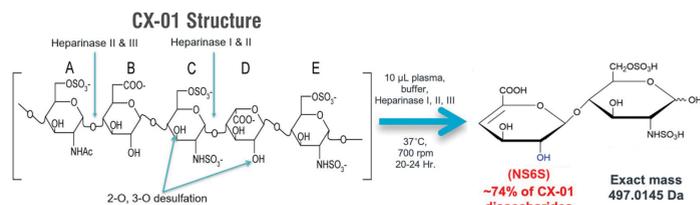
- Identification of a unique Enoxaparin disaccharide (NS2S) hydrolysis product
- Derivation of NS6S-to-NS2S response relationship over the relevant Enoxaparin range
- Compensation for non-equivalent Enoxaparin hydrolysis and derivatization efficiencies under CX-01 optimized conditions
 - Run specific curve fitting

METHOD

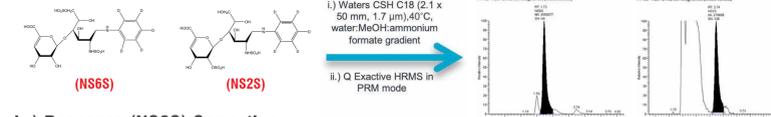
The method described quantifies CX-01 in K₂EDTA human plasma over a target assay range of 0.0500 – 50.0 µg/mL in the presence of concomitantly dosed Enoxaparin. The effective CX-01 concentration range is determined on a run-by-run basis and by the measured quality of signature disaccharide (NS6S) response correction near the low end of the assay by including in-run concomitantly spiked QC samples. The method is based on the enzymatic hydrolysis of CX-01 and Enoxaparin in human plasma to generate signature disaccharides for the primary and concomitant analytes of interest, the addition of stable label ISTD disaccharides, protein precipitation of the hydrolysis mixture, derivatization^{2,3} of signature disaccharides and ISTDs, and LC-HRMS in PRM mode for instrumental analysis. Subsequent response correction for the concomitantly dosed Enoxaparin is conducted as described herein to arrive at an accurate target therapeutic (CX-01) concentration determination.

METHODOLOGY OVERVIEW:

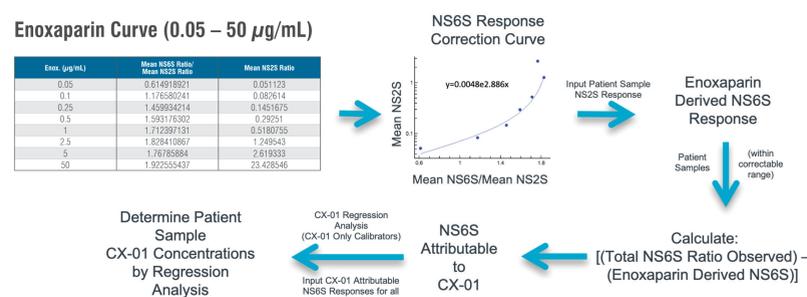
i.) Sample Hydrolysis



iii.) Separation and Detection



iv.) Response (NS6S) Correction



RESULTS

The method was qualified in a single run. Assay performance demonstrated individual calibration standard %biases ranging from -6.0% - +6.0% for the 8-point CX-01 calibration line assayed in duplicate (Table 1). A mean %bias of -2.0% to +0.5% was observed across four levels of CX-01 QCs (Table 2). Three additional QCs, designed to characterize individual NS6S response correction, were prepared at low (0.150 µg/mL) and high (40.0 µg/mL) CX-01 concentrations to also contain 0.500 µg/mL, 2.00 µg/mL, or 10.0 µg/mL Enoxaparin (Table 3). These additional QCs demonstrated NS6S response corrected %biases from -1.3% to -13.3% for samples containing up to 2.00 µg/mL Enoxaparin. The low QC %bias exceeded correctability tolerances at the 10.0 µg/mL Enoxaparin level. For the high QC samples, %bias ranged from +3.5% to +5.5% across the concentrations of Enoxaparin tested.

Additionally, an eight-point CX-01 calibration line spanning the target assay range (0.0500 – 50.0 µg/mL) and containing an ~Cmax Enoxaparin level (5 µg/mL), was assayed and subjected to in-run NS6S response correction. Corrected NS6S response values for individual combinations spiked calibration standards demonstrated 87% – 103% of the observed response from CX-01 - only spiked calibrators prepared at the same nominal levels over a CX-01 range of 0.25 -50 µg/mL (Table 4). Corrected response values across the two lowest CX-01 calibrator levels (0.0500 – 0.100 µg/mL) ranged from -137% to +169%, demonstrating an expected limitation to the response correction applied.

Additional method qualification parameters assessed included room temperature Enoxaparin benchtop stability in plasma and potential for variability in assay performance when processing Enoxaparin from various sources/vendors. The ~25-hour benchtop stability samples demonstrated responses ranging from 89.4% to 103.2% of fresh preparations (Table 5). Observed NS6S ratios from Enoxaparin supplied by four different vendors varied from +105% to +153% of the mean observed ratios from a US Pharmacopeia standard (Table 6).

Three runs were conducted for non-regulated sample analysis of 66 phases 2/3 clinical samples in patients with severe COVID-19 using the qualified method that met the acceptance criteria for passing runs. Incurred sample reproducibility (ISR) was also assessed by the assay of at least 10% of the study samples. All paired original and repeat assay results for the ten samples undergoing ISR analysis were within the acceptable limits (+/- 20.0%) for CX-01.

Method Qualification and Phase 2/3 Study ISR Results:

Table 1. Method Qualification Run - CX-01 Calibration line Results

Qual. Curve Number	CX-01 0.050 µg/mL	CX-01 0.100 µg/mL	CX-01 0.250 µg/mL	CX-01 0.500 µg/mL	CX-01 1.000 µg/mL	CX-01 2.500 µg/mL	CX-01 5.000 µg/mL	CX-01 50.0 µg/mL	%Bias
5	0.0500	0.1000	0.2500	0.5000	1.0000	2.5000	5.0000	50.2	-1.1

Table 2. Method Qualification Run - CX-01 QC/VS Results

Curve Number	LOW VS 0.150 µg/mL CX-01	%Bias	LOW VS 2.00 µg/mL Enoxaparin	%Bias	MIDDLE VS 10.0 µg/mL CX-01	%Bias	HIGH VS 40.0 µg/mL CX-01	%Bias
5	0.0988	-2.9	0.992	-0.8	5.08	1.6	40.8	-1.3

Table 3. Method Qualification Run - CX-01 + Enoxaparin Response Correction QC/VS Results

Curve Number	LOW VS 0.150 µg/mL CX-01	%Bias	LOW VS 2.00 µg/mL Enoxaparin	%Bias	LOW VS 10.0 µg/mL CX-01	%Bias	HIGH VS 40.0 µg/mL CX-01	%Bias	HIGH VS 2.00 µg/mL Enoxaparin	%Bias	HIGH VS 10.0 µg/mL CX-01	%Bias
5	0.149	-0.7	0.118	-21.3	0.196	-9.3	41	2.5	42.5	6.3	41.7	4.3

RESULTS (CONTINUED)

Table 4. Method Qualification Run - CX-01 + ~Cmax Enoxaparin – Correction Results

Calibration Standard (CS) CX-01 (Enoxaparin) µg/mL	Sample Name	Total NS6S Peak Area Ratio Detected (CX-01 + Enoxaparin)	Enoxaparin Attributable NS6S Peak Area Ratio (via NS2S Derivation)	CX-01 Attributable NS6S Corrected Peak Area Ratio	CX-01 Only Spiked CS NS6S Peak Area Ratio	NS6S Correction Performance (Corrected NS6S Ratio from Calibrated CS vs % Observed NS6S Ratio from CX-01 only CS)
0.0500 (0.00)	ES CX01 0.05 1	4.840244	4.8984078	0.2541362	0.226025	169%
0.100 (0.00)	ES CX01 0.1 1	5.101162	0.78314372	0.455963	0.455963	99%
0.250 (0.00)	ES CX01 0.25 1	5.646511	4.46807229	1.17843371	1.192193	87%
0.500 (0.00)	ES CX01 0.5 1	6.735343	4.726180079	2.02916221	2.321934	98%
1.00 (0.00)	ES CX01 1 1	9.258416	4.59246994	4.64193208	4.758301	89%
2.50 (0.00)	ES CX01 2.5 1	15.425192	4.720178338	10.7051356	12.00807	90%
5.00 (0.00)	ES CX01 5 1	27.098882	4.63846279	22.4603572	24.835572	100%
10.0 (0.00)	ES CX01 10 1	24.416563	4.465841734	236.9507853	233.029163	100%
50.0 (0.00)	ES CX01 50 2	244.670276	4.268982916	246.2728911	236.261466	100%
5.00 (0.00)	ES CX01 5 2	27.501933	4.767504469	22.73442833	23.563356	96%
2.50 (0.00)	ES CX01 2.5 2	16.099827	4.83878542	11.26394846	11.049215	102%
1.00 (0.00)	ES CX01 1 2	8.716138	4.55588276	4.20280734	4.546538	93%
0.500 (0.00)	ES CX01 0.5 2	6.985195	4.71577189	2.269417811	2.246951	101%
0.250 (0.00)	ES CX01 0.25 2	5.656462	4.45157086	1.20489114	1.172627	103%
0.100 (0.00)	ES CX01 0.1 2	4.917285	4.863703902	0.033891096	0.42368	8%
0.0500 (0.00)	ES CX01 0.05 2	4.837628	4.93765828	-0.303318028	0.219513	-137%

Table 5. Method Qualification Run - Enoxaparin Benchtop Stability Results

Sample Name	Enoxaparin (µg/mL)	NS6S Peak Area Ratio	Mean NS6S Peak Area Ratio (per supplier and level)	NS6S Ratio as % of Mean USP Material
BT 24Hr Enox 0.1 1	0.100	0.091588	-	99.2%
BT 24Hr Enox 0.1 2	0.100	0.091258	-	89.4%
BT 24Hr Enox 1 1	1.00	0.915888	-	103.2%
BT 24Hr Enox 1 2	1.00	0.91131	-	102.7%
BT 24Hr Enox 5 1	5.00	4.453883	-	94.6%
BT 24Hr Enox 5 2	5.00	4.1406	-	93.9%
USP Enoxaparin 0.1 1	0.100	0.099676	0.099702	-
USP Enoxaparin 0.1 2	0.100	0.891151	0.891151	-
USP Enoxaparin 5 1	5.00	4.750316	4.630611	-
USP Enoxaparin 0.1 2	0.100	0.094578	-	-
USP Enoxaparin 1 2	1.00	5.28982*	-	-
USP Enoxaparin 5 2	5.00	4.510306	-	-

Table 6. Method Qualification Run - Enoxaparin Source/Vendor Results

Sample Name	Enoxaparin (µg/mL)	NS6S Peak Area Ratio	Mean NS6S Peak Area Ratio (per supplier and level)	NS6S Ratio as % of USP Standard
Amphastar ONLY 0.1 1	0.100	0.131748	0.1321675	136%
Amphastar ONLY 1 1	1.00	1.102758	1.1424265	129%
Amphastar ONLY 5 1	5.00	5.812325	5.7029395	123%
Amphastar ONLY 5 2	5.00	5.586704	-	-
Amphastar ONLY 1 2	1.00	1.182115	-	-
Amphastar ONLY 0.1 2	0.100	0.12282	-	-
Winthrop ONLY 0.1 1	0.100	0.13243	0.1377545	142%
Winthrop ONLY 1 1	1.00	1.31801	1.355812	153%
Winthrop ONLY 5 1	5.00	5.30145	5.377683	116%
Winthrop ONLY 1 2	1.00	5.433016	-	-
Winthrop ONLY 1 2	1.00	1.392623	-	-
Winthrop ONLY 0.1 1	0.100	0.10129	-	-
Lexvex ONLY 1 1	1.00	0.95193	0.97016	109%
Lexvex ONLY 5 1	5.00	4.85046	4.845246	105%
Lexvex ONLY 5 2	5.00	4.84046	-	-
Lexvex ONLY 1 2	1.00	0.98807	-	-
Lexvex ONLY 0.1 2	0.100	0.107883	-	-
USP Enoxaparin 0.1 1	0.100	0.099676	0.099702	-
USP Enoxaparin 1 1	1.00	0.891151	0.891151	-
USP Enoxaparin 5 1	5.00	4.750316	4.630611	-
USP Enoxaparin 0.1 2	0.100	0.094578	-	-
USP Enoxaparin 1 2	1.00	5.28982*	-	-
USP Enoxaparin 5 2	5.00	4.510306	-	-

Table 7. Phase 2/3 Study ISR Results

Water ID	Custom ID	Sample ID	Group	Subject	Treatment	Time Test	Final Analyte	Final Original Concentration (µg/mL)	ISR Run ID	ISR Concentration (µg/mL)	% Difference	Flag
11	6511803544-02	6511803544-02 0001-002 A Plasma-1 Day 4 / Day 4	1	0001-002	A	Day 4	3	3.03	6	2.65	-13.4	
34	6511496004-06	6511496004-06 0001-005 A Plasma-1 Early Therapy Discontinuation / Day 988	1	0001-005	A	Day 4	3	3.15	6	2.96	-6.22	
48	6511803599-06	6511803599-06 0001-007 A Plasma-1 Early Therapy Discontinuation / Day 988	1	0001-007	A	Day 4	3	5.82	6	5.92	1.7	
74	6511803593-02	6511803593-02 0002-002 A Plasma-1 Day 4 / Day 4	1	0002-002	A	Day 4	3	2.08	6	1.91	-8.52	
95	6511803591-02	6511803591-02 0002-005 A Plasma-1 Day 4 / Day 4	1	0002-005	A	Day 4	3	3.06	6	3.16	3.22	
96	6511803596-02	6511803596-02 0002-005 A Plasma-1 Day 8 / Day 8	1	0002-005	A	Day 6	3	3.47	6	3.51	1.15	
108	6511803636-02	6511803636-02 0003-003 A Plasma-1 Day 2 / Day 2	1	0003-003	A	Day 2	3	1.64	6	1.59	-3.1	
115	6512027390-02	6512027390-02 0007-001 A Plasma-1 Day 2 / Day 2	1	0007-001	A	Day 2	4	2.35	6	2.56	8.55	
116	6512027395-02	6512027395-02 0007-001 A Plasma-1 Day 4 / Day 4	1	0007-001	A	Day 4	4	2.12	6	2.15	1.41	
157	6511803638-02	6511803638-02 0015-001 A Plasma-1 Day 2 / Day 2	1	0015-001	A	Day 2	4	4.75	6	4.35	-8.19	

CONCLUSIONS

CX-01 quantitation in the presence of concomitantly dosed Enoxaparin requires an in-run signature disaccharide response correction approach. The assay requires the determination of concomitant medication concentration via alternate disaccharide monitoring and subsequent correction of the total signature disaccharide response detected to arrive at an accurate target therapeutic concentration determination. Run-specific curve fitting, relative to co-dosed QC samples intended to mimic patient samples containing up to Cmax levels of Enoxaparin, as well as ISR results from a Phase 2/3 study for which the method was deployed, demonstrated reasonable response correction and highly reproducible CX-01 concentration determinations from concomitantly-dosed patient samples.

REFERENCES

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